## EFFECT OF PRIMING ON GERMINATION AND INITIAL GROWTH OF MELON PLANTS UNDER SALT STRESS

CASTAÑARES, J. L.<sup>1</sup> & BOUZO, C. A.<sup>2</sup>

#### **ABSTRACT**

The objective of this work was to evaluate the effect of different osmotic agents and priming durations on the germination and initial growth of melon in salinity. The osmotic agents were NaCl and CaCl<sub>2</sub> and durations 2 and 4 days. First a response curve of germination to salinity was made. After priming the seeds were placed to germinate in a saline medium (14 dS m<sup>-1</sup>). The germination percentage was determined. The two treatments with the best results (NaCl and CaCl<sub>2</sub> 2 days in both salts) were selected to evaluate the initial growth of the plants in salinity. It was sown in pots, irrigated with nutrient solution and 60 mM NaCl. The germination was reduced to 50% with 14 dS m<sup>-1</sup> and inhibited with 22 dS m<sup>-1</sup>. The best germination was achieved with NaCl and CaCl<sub>2</sub> 2 days with 99 and 94% respectively. Both treatments showed an increase in growth parameters and chlorophyll content.

Key words: priming, germination, salinity, stress.

#### RESUMEN

## Efecto del priming en la germinación y crecimiento inicial de plantas de melón en estrés salino.

El objetivo del trabajo fue evaluar el efecto de diferentes agentes osmóticos y tiempos de *priming* en la germinación y crecimiento inicial de melón en salinidad. Los agentes osmóticos fueron NaCl y CaCl<sub>2</sub> y los tiempos del tratamiento 2 y 4 días. Primeramente se confeccionó una curva de respuesta de la germinación a la salinidad. Luego del *priming* las semillas se colocaron a germinar en

<sup>2.-</sup> Departamento de Ciencias Básicas. Universidad Nacional de Luján. Rutas NAcionales 5 y 7. (6700) Luján provincia de Buenos Aires. Email: joseluis.cas@gmail.com

un medio salino (14 dS m<sup>-1</sup>). Se determinó el porcentaje de germinación. Se seleccionaron los dos tratamientos con los mejores resultados (NaCl y CaCl<sub>2</sub> 2 días en ambas sales) para evaluar el crecimiento inicial de las plantas en salinidad. Se sembró en macetas, regándose con solución nutritiva y 60 mM de NaCl. La germinación se redujo al 50% con 14 dS m<sup>-1</sup> y se inhibió a 22 dS m<sup>-1</sup>. La mejor germinación se logró con NaCl y CaCl<sub>2</sub> a los 2 días con 99 y 94% respectivamente. Ambos tratamientos mostraron un incremento de los parámetros de crecimiento y clorofila.

Palabras clave: priming, germinación, salinidad, estrés.

#### INTRODUCTION

Soil salinity is one of the most important conditions for growing melon. Salinity can conduce to a reduction of percentage and germination rate (6; 46). In addition, in severe salt stress conditions, the seedlings can show a reduction of aerial biomass (28), inhibition of stem, leaf and root growth (5), reduction of leaf water potential and stomatal conductance (29), metabolic alterations with photosynthesis decrease (14) and reduced fruit yield and quality (32; 14). However, if development of plants is considered, germination and initial seedling growth phases have the highest sensitivity to salinity, increasing tolerance between fruit development and harvest (37; 17; 9; 45). The technique of pre-conditioning seeds (priming) can become a technological alternative to overcome limitations imposed by salinity.

Priming is a controlled seed hydration technique that allows activating the metabolic and repair processes (activation of DNA repair pathways and antioxidant mechanisms) that occur in early stages of imbibition (pre-germinative metabolism),

without radicle emission (germination) (38). Priming treatments have proven their advantages and are investigated for the following purposes: a) seed invigoration during storage; b) increase, acceleration and synchronization of germination and establishment of seedlings; c) overcoming or alleviation of different types of dormancy and; d) improvement of germination and establishment of seedlings under adverse environmental conditions (25; 41).

Controlled hydration can be achieved with an osmotic agent that reduces the osmotic potential (Ψo) of water (osmopriming), a substance that retains surface water (matric priming) or the addition of a limited amount of water (hydropriming) (49). Osmopriming is the main technique used because of simplicity and satisfactory results (22).

The beneficial effects of priming are not exclusive of seed germination and can be can be extended to later stages by increasing tolerance to different biotic and abiotic stresses (11; 13; 35).

The objective of this work was to evaluate the effect of different osmotic agents and priming durations on germination and initial growth of melon plants under salinity stress.

#### **MATERIALS AND METHODS**

Seeds of Honeydew melon (*Cucumis melo* L.) were used in three experiments to determine the following effects: 1) salinity stress in germination; 2) priming on seed germination in salinity; and 3) priming in the initial growth and chlorophyll content of melon plants in salinity.

## Experiment 1: Effect of salinity on germination

Seeds were placed between two filter papers (Whatman no 1), in 8.5 cm diameter Petri dishes, soaked with different NaCl concentrations (0 to 22 dS m<sup>-1</sup>) in a ratio 1: 5 (weight: volume) (15). Ten seeds per dish were used, equidistant from each other, with 5 replicates per concentration. The assay was performed in a germination chamber (Semedic I-500 PF) at 25 °C (± 0.5 °C) and 1000 lux of light intensity. After 8 days germination was recorded (23).

### Experiment 2: Effect of priming on germination in salinity conditions

Priming was performed in culture chamber in dark and at 25 °C (± 0.5 °C). The seeds were positioned equidistant from each other on a filter paper (Whatman no 1) in 8.5 cm diameter Petri dishes, soaked with two different osmotic solutions: NaCl and CaCl2, in a ratio of 1: 5 (weight: volume). The concentration of these osmotic agents was calculated to achieve an osmotic potential (Ψo) of -1.5 MPa according to the Van't Hoff equation for Ψo (44). Two treatment durations were studied: 2 and 4 days. After priming the seeds were removed and rinsed three times in distilled water. Later the seeds were put to germinate in the same way as in the previous experiment, imbibed with the corresponding saline solution that caused a 50% reduction of germination.

Control treatment consisted in sowing seeds without priming treatment. After 8 days the number of germinated seeds was recorded and germination percentage (GP) was calculated (23). Five replicates were used with 10 seeds per treatment. A completely randomized design was used. The data obtained from the experiment were analyzed using an analysis of variance (ANOVA).

# Experiment 3: Effect of priming on initial growth of melon plants under salinity conditions

The two treatments of the previous experiment in which the best response in germination was recorded were chosen.

Sowing was done in 1000 cm<sup>3</sup> pots, previously filled with perlite. Irrigation was performed daily with the Hoagland's nutrient solution (21). After first leaf emission, plants were divided into two groups: a) irrigated with nutritive solution with 2.0 dS m<sup>-1</sup> electrical conductivity (EC) (S0); and b) irrigated with nutritive solution with the addition of 60 mM NaCl, to achieve an EC of 8.0 dS m<sup>-1</sup> (S1). The EC was monitored daily with a Milwaukee conductivity meter, model WP MC66 0-10MS/C.

The experiment was performed in a phytotron, with a 16 h light/8 h darkness cycle (3000 lux light intensity) and temperatures of 25/18 °C ( $\pm$  1.0 °C), considered as optimal for the growth of this specie (10; 30). Five plants were extracted from each treatment every 10 days and the following parameters were determined: total dry weight, main stem length, number of leaves, leaf area from digitalized images of leaves using the Image J © computer program, and total leaf chlorophyll by spectrophotometry (33). 50 plants per treatment were used in a completely randomized design. Data measured during the experiment were statistically analyzed using ANOVA.

#### **RESULTS**

#### Experiment 1

Reduction in germination was gradual with increasing EC. Values between 14 and 16 dS m<sup>-1</sup> reduced germination to 50%, while it was completely inhibited at 22 dS m<sup>-1</sup> (Figure 1).

#### Experiment 2

Priming partially reversed the inhibitory effect of salinity. However the best results were obtained, for both salts, with 2 days duration (Table 1).

It was observed that although priming enhanced germination percentage with both salts, increasing priming duration, from 2 to 4 days, decreased the favorable treatment effect.

#### Experiment 3

This experiment, in which the effect of salinity on melon plants was studied, was performed with the two treatments in which the best germination percentage was recorded (NaCl and CaCl,, and 2 days). The effect of these treatments on growth parameters is represented in figure 2. High EC had a negative effect on total dry weight, number of leaves, leaf area and main stem length. In the measured variables, priming allowed increasing salinity tolerance, with a positive differentiation respect to control. However, in low salinity condition (S0) there were no differences between the treatments and the control, therefore the treatments with priming did not present increases in the measured variables (Figure 2).

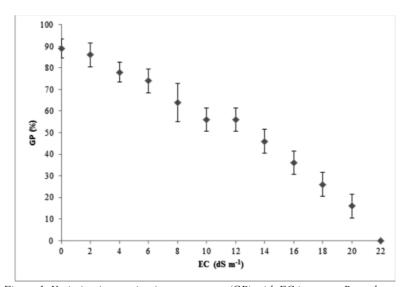


Figure 1. Variation in germination percentage (GP) with EC increase. Bars show  $\pm$  SE.

Table 1. Germination percentage at 14 dS m<sup>-1</sup> with different priming variations.

Priming	GP
NaCl 2.days	99.0 a
CaCl <sub>2</sub> 2.days	94.0 a
NaCl 4.days	74.4 b
CaCl <sub>2</sub> 4.days	66.2 b
Control	50.2 c

Different letters indicate a significative difference ( $p \le 0.05$ ) according to Tukey.

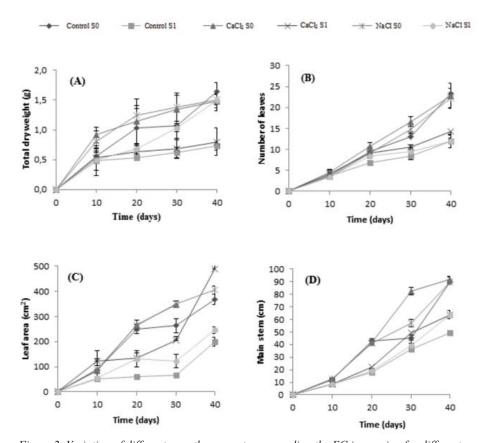


Figure 2. Variation of different growth parameters according the EC increasing for different priming variations: dry weight (A), leaves number (B), leaf area (C), and length of main stem (D). Control S0: Control treatment with EC of nutrient solution 2 dS  $m^{-1}$ ; Control S1: Control treatment with EC of nutrient solution 8 dS  $m^{-1}$ ; CaCl<sub>2</sub>S0: priming with CaCl<sub>2</sub> and EC of nutrient solution 2 dS  $m^{-1}$ ; CaCl<sub>2</sub>S1: priming with CaCl<sub>2</sub> and EC of nutrient solution 8 dS  $m^{-1}$ ; NaCl S0: priming with NaCl and EC of nutrient solution 2 dS  $m^{-1}$ ; NaCl S1: priming with NaCl and EC of the nutrient solution 8 dS  $m^{-1}$ . Bars show  $\pm$  SE.

#### J. L. Castañares et al.

The concentration of total leaf chlorophyll after 40 days of sowing showed a reduction due to salinity, although a higher concentration was measured in plants coming from seeds with priming treatment, both in low (S0) and high (S1) salinity conditions (Figure 3).

#### DISCUSSION

Ψo of -1.5 MPa was used because lower Ψo (< -2.0 MPa) could be harmful to seeds (7) and Ψo close to 0 MPa could allow germination during priming. The above considerations are further justified since for this species Ψo of -1.3 MPa has been determined as the most suitable for safe priming (36).

From the point of view of osmotic agents, polyethyleneglycol (PEG) has traditionally been used for priming, since it's an inert compound and because of the relatively high molecule size it reduces the Ψo of the solution without absorption of the molecule into the seed (50). However, this compound has an extremely high viscosity which limits oxygen transfer within the solution, as well as the subsequent removal from seeds (38). For this reason, the use of inorganic salts could overcome these constraints, in addition to reducing costs (20; 31). The results obtained confirm the beneficial effect of NaCl and CaCl, as osmotic agents.

For these salts, their use has been studied for different species (38). In case of NaCl, Farhoudi *et al.* (16), working with canola (*Brassica napus* L.), observed that the po-

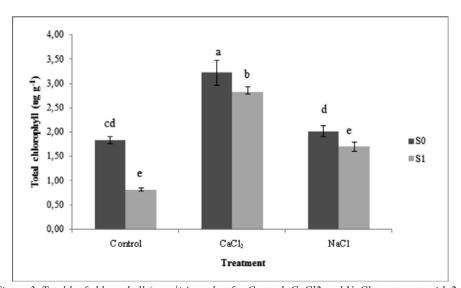


Figure 3. Total leaf chlorophyll ( $\mu g g^1$ ) in melon for Control, CaCl2 and NaCl treatments with 2 (S0) and 8 (S1) dS  $m^1$ , measured 40 days after transplant. Different letters on the bars indicate a significative difference ( $p \le 0.05$ ) according to Tukey. Vertical bars show  $\pm$  SE.

sitive response of priming to germination and initial plant growth could be associated with the activation of antioxidant systems and proline accumulation. This is consistent with that observed by Aloui et al. (1), who worked with pepper (Capsicum annuum L.). Passam and Kakourioitis (39) reported an increase in germination in such conditions, although the effects would not extend beyond this initial state. Nakaune et al. (34) proposed that the effect of priming with NaCl in tomato (Solanum lycopersicum L.) would be related to an increase of gibberellin content, a hormone linked to germination, as a consequence of the activation of genes associated with its synthesis. Also in this specie it was detected a higher Na<sup>+</sup> and Cl<sup>-</sup> content in seeds and roots, which conduces to a greater capacity of osmotic adjustment (11).

In relation to CaCl,, in bean (Cajanus cajan L.), a semi-perennial shrub legume, Jyotsna and Srivastava (26), observed that seedlings from primed seeds, when growing in saline soil, showed an increase in their protein composition and soluble sugars. In addition to the own advantages of a seed with a controlled start of its germination provided by priming, possibly the use of CaCl, in saline conditions could be associated to the influence of Ca<sup>2+</sup> on the membrane integrity, as well as the antioxidant effect of several enzymes in which participates as cofactor (19; 51). In maize (Zea mays L.) growing in salinity it was observed that, when comparing different chlorine salts as osmotics agents, with CaCl, it was registered a higher content of Ca<sup>2+</sup> in seeds, mesocotyl and radicle (4).

The lower germination measured when priming duration increased (Table 1) can be explained by the fact that an extensive partial imbibition, without the radicle emission, could cause a depletion of seminal reserves. This situation leads to a reduction in seed vigor and the consequent reduction of germination under unfavorable conditions (8; 27) as well as irregular water absorption with the consequent loss of vital electrolytes for seeds (24).

The improvement in growth parameters in plants from primed seeds (Figure 2), evidences that the advantages of priming can extend beyond the seed state, phenomenon known as "priming memory" (12), which results in a better adaptive capacity for the saline stress situation.

The measured reduction in chlorophyll concentration in plants growing in saline conditions (Figure 3) is considered as a result of oxidative stress (47) and would be associated with the presence of ions of the salts in some stages of the chlorophyll synthesis, causing the reduction of this molecule (2). However the activation of degradation mechanisms mediated by the chlorophyllase enzyme (42; 48) cannot be ruled out either. Priming allowed increasing the chlorophyll concentration (Figure 3). Some antecedents that explain this effect are related to the fact that priming allows to reverse partially the damage caused by oxidative stress (3; 40; 43), evidenced in the increase of chlorophyll concentration.

#### **CONCLUSIONS**

Priming with NaCl and CaCl<sub>2</sub> allowed increasing the germination percentage, growth's parameters and chlorophyll contents of plants in salinity stress. The 2 days duration of treatment results more adequate for the studied conditions.

#### **BIBLIOGRAPHY**

- 1.- ALOUI, H.; SOUGUIR, M.; LATIQUE, S.; HANNACHI, C. 2014. Germination and growth control and primed seeds of pepper as affected by salt stress. Cercetari agronomice în Moldova, 47: 83-95.
- 2.- ALI, Y.; ASLAM, Z.; ASHRAF, M.; TAHIR, G. R. 2004. Effect of salinity on chlorophyll concentration, leaf area, yield and yield components of rice genotypes grown under saline environment. International Journal of Environmental Science and Technology 1: 221-225.
- 3.- ANSARI, O.; SHARIF-ZADEH, F. 2012. Osmo and hydro priming improvement germination characteristics and enzyme activity of Mountain Rye (*Secale montanum*) seeds under drought stress. Journal of Stress Physiological Biochemistry 8: 253-261.
- 4.- ASHRAF, M.; RAUF, H. 2001. Inducing salt tolerance in maize (*Zea mays* L.) through seed priming with chloride salts: Growth and ion transport at early growth stages. Acta Physiologiae Plantarum, 23:407-414.
- 5.- BOROCHOV-NEORI, H.; SHANI, U. 1995. Response of melon plants to salt.
  2. Modulation by root growth temperature the role of root membrane properties.
  Journal of Plant Physiology 145: 545-550.
- 6.- BOTÍA, P.; CARVAJAL, M.; CER-DÁ, A.; MARTÍNEZ. 1998. Response of eight *Cucumis melo* cultivars to salinity during germination and early vegetative growth. Agronomie 18: 503-513.
- 7.- BRADFORD, K. J. 1986. Manipulation of seed water relations via osmotic priming improves germination under stress conditions. Seed germination under environmental stress. HortScience 2: 1105-1112.
- 8.- BRADFORD, K. J.; STILL, D. W. 2004. Application of hydrotime analysis in seed testing. Seed Technology 26:75-85.

- 9.- CARVAJAL, M.; DEL AMOR, F.M.; FER-NÁNDEZ-BALLESTER, G.; MARTÍNEZ, V.; ANTONIO CERDÁ, A. 1998. Time course of solute accumulation and water relations in muskmelon plants exposed to salt during different growth stages. Plant Science 138: 103-112.
- 10.- CASACA, A. D. 2005. Guías tecnológicas de frutas y vegetales. Número 12: El Melón. Proyecto de Modernización de los Servicios de Tecnología Agrícola, PROMSTA. El Salvador. 13p.
- 11.- CAYUELA, E.; PEREZ-ALFOCEA, F.; CARO, M.; BOLARÍN, M.C. 1996. Priming of seeds with NaCl induces physiological changes in tomato plants grown under salt stress. Physiologic Plantarum 96: 231-236.
- 12.- CHEN, K.; ARORA, R. 2013. Priming memory invokes seed stress-tolerance. Environmental and Experimental Botany 94: 33-45.
- 13.- CUARTERO, J.; FERNÁNDEZ-MU-ÑOZ, R. 1999. Tomato and Salinity. Scientia Horticulturae 78: 83-125.
- 14.- DEL AMOR, F.M.; RUIZ-SÁNCHEZ, M.C.; MARTÍNEZ, V.; CERDA, A. 2000. Gas exchange, water relations, and ion concentrations of salt-stressed tomato and melon plants. Journal of Plant Nutrition 23: 1315-1325.
- **15.- FAROOQ, M.; IRFAN, M.; AZIZ, T.; AHMAD, I.; CHEEMA, S.** 2013. Seed priming with ascorbic acid improves drought resistance of wheat. Journal of Agronomy and Crop Science 199 (1): 12-22.
- 16.- FARHOUDI, R.; SHARIFZADEH, F.; POUSTINI, K.; MAKKIZADEH, M.T.; KOCHAK POR, M. 2007. The effects of NaCl priming on salt tolerance in canola (*Brassica napus*) seedlings grown under saline conditions. Seed Science and Technology 35 (3): 754-759.
- 17.- FRANCO J.A.; ESTEBAN, C.; RODRÍ-GUEZ, C. 1993. Effects of salinity on various growth stages of muskmelon cv. Revigal. Journal of Horticultural Science 68: 899-904.

- **18.- GUZMÁN, M.; OLAVE, J.** 2006. Response of growth and biomass production of primed melon seed (*Cucumis melo* L. cv. Primal) to germination salinity level and N-forms in nursery. Journal of Food, Agriculture and Environment 4: 163-165.
- 19.- HAMEED, A.; AFZAL, I.; IQBAL, N. 2010. Seed priming and salinity induced variations in wheat (*Triticum aestivum* L.) leaf protein profile. Seed Science and Technology 38: 236-241.
- 20.- HAN YOON, B. Y.; LANG, H. J.; COBB, B. G. 1997. Priming with salt solutions improves germination of pansy seed at high temperatures. HortScience 32 (2): 248-250.
- HOAGLAND, D. R.; ARNON, D. I. 1950. The water-culture method for growing plants without soil. Circular California Agricultural Experiment Station, 347.
- **22.- IBRAHIM, E. A.** 2016. Seed priming to alleviate salinity stress in germinating seeds. Journal of Plant Physiology 192: 38-46.
- 23.- INTERNATIONAL SEED TESTING ASSOCIATION (ISTA). 2009. International Rules for Seed Testing. Seed Science and Technology 27: 1-333.
- 24.- JET, L. W.; EELBAUM, G. R.; MOR-SE, R. D. 1996. Effect of matric and osmotic priming treatments on broccoli seed germination. Journal of American Society of Horticultural Science 121:423-429.
- 25.- JISHA, K. C.; VIJAYAKUMARI, K.; PUTHUR, J. T. 2013. Seed priming for abiotic stress tolerance: an overview. Acta Physiologiae Plantarum 35:1381-1396.
- 26.- JYOTSNA, V.; SRIVASTAVA, A. K. 1998. Physiological basis of salt stress resistance in pigeon pea (*Cajanus cajan* L.). II. Pre-sowing seed soaking treatment in regulating early seedling metabolism during seed germination. Plant Physiological Biochemistry 25: 89-94.

- 27.- KRISHNAN P.; NAGARAJAN, S.; MO-HARIR, A. V. 2004. Thermodynamic Characterisation of Seed Deterioration during Storage under Accelerated Ageing Conditions. Biosystems Engineering 89: 425-433.
- 28.- KUSVURAN, S.; YASAR, F.; ELLIATIO-GLU, S.; ABAK, K. 2007. Utilizing some of screening methods in order to determinate of tolerance of salt stress in the melon (*Cucumis melo* L.). Research Journal of Agricultural and Biological Sciences 3: 40-45.
- 29.- KUSVURAN, S. 2012. Effects of drought and salt stresses on growth, stomatal conductance, leaf water and osmotic potentials of melon genotypes (*Cucumis melo L.*). African Journal of Agricultural Research 7: 775-781.
- 30.- MÁRMOL, J. 2008. Cultivo de melón en invernadero. Junta de Andalucía, Consejería de Agricultura y Pesca. Madrid. España. 60p.
- 31.- MAUROMICALE, G.; CAVALLARO, V. 1997. A comparative study of different compounds on priming of tomato seed germination under suboptimal temperatures. Seed Science Technology 25: 399-408.
- **32.- MENDLINGER, S.** 1994. Effect of increasing plant density and salinity on yield and fruit quality in muskmelon. Scientia Horticulturae 57: 41-49.
- **33.- MOSS, B.** 1967. A spectrophotometric method for estimation of percentage degradation of chlorophylls to phaeo-pigments extract of algae. Limnology Oceanography 12: 335-340.
- 34.- NAKAUNE, M.; HANADA, A.; YIN, Y. G.; MATSUKURA, C.; YAMAGU-CHI, S. 2012. Molecular and physiological dissection of enhanced seed germination using short-term low-concentration salt seed priming in tomato. Plant Physiological Biotechnology 52: 28-37.
- 35.- NASCIMENTO, W. M. 2002. Sementes de melãoosmoticamentecondicionadas: valeapena utilizálas? Horticultura Brasileira 20: 133-135.

- **36.- NASCIMENTO, W. M.** 2003. Muskmelon seed germination and seedling development in response to seed priming. Scientia Agricola 60: 71-75.
- 37.- NUKAYA A.; MASUI, M.; ISHIDA, A. 1984. Salt tolerance of muskmelon as affected by dilute sea water applied at different growth stages in nutrient solution culture. Journal of the Japanese Society for Horticultural Science 53: 168-175.
- 38.- PAPARELLA, S.; ARAUJO, S. S.; ROS-SI, G.; WIJAYASINGHE, M.; CARBO-NERA, D.; BALESTRAZZI, A. 2015. Seed priming: state of the art and new perspectives. Plant Cell Reports 34(8): 1281-1293.
- 39.- PASSAM, H. C.; KARAVITES, P. I.; PAPANDREOU, A.; THANOS, C. A.; GEORGHIOU, K. 1989. Osmoconditioning of seeds in relation to growth and fruit yield of aubergine, pepper, cucumber and melon in unheated greenhouse cultivation. Scientia Horticulturae 38: 217-216.
- **40.- REJEB, K.B.; ABDELLY, C.; SAVOU-RE, A.** 2014. How reactive oxygen species and proline face stress together. Plant Physiological. Biochemistry 80: 278-284.
- 41.- SÁNCHEZ, J. A.; ORTA, R.; MUÑOZ, B. C. 2001. Tratamientos pregerminativos de hidratación-deshidratación de las semillas y sus efectos en plantas de interés agrícola. Agronomía Costarricense 25: 67-91.
- 42.- SANTOS, C.V. 2004. Regulation of chlorophyll biosynthesis and degradation by salt stress in sunflower leaves. Scientia Horticulturae 103: 93-99.
- 43.- SEDGHI, M.; AMANPOUR-BALA-NEJI, B.; BAKHSHI, J. 2014. Physiological enhancement of medicinal pumpkin seeds (*Cucurbita pepo var. styriaca*) with different priming methods. Irani Journal of Plant Physiology 5: 1209-1215.

- 44.- SHABALA, S.; MUNNS, R. 2012. Salinity stress: physiological constraints and adaptative mechanism. En: Shabala, S. (Ed.). Plant stress physiology. CAB international. Londres, Inglaterra. p. 59-93.
- 45.- SIVRITEPE N.; SIVRITEPE, H. O.; ERIS, A. 2003. The effects of NaCl priming on salt tolerance in melon seedlings grown under saline conditions. Scientia Horticulturae 97: 229-237.
- 46.- SIVRITEPE, H.; SIVRITEPE, N.; ERIS, A.; TURHAN, E. 2005. The effects of NaCl pre-treatments on salt tolerance of melons grown under long-term salinity. Scientia Horticulturae 106: 568-581.
- **47.- SMIRNOFF, N.** 1996. Botanical briefing: the function and metabolism of ascorbic acid in plants. Annals of Botany 78: 661-669.
- 48.- TAÏBI, K.; TAÏBI, F.; ABDERRAHIMA, L. A.; ENNAJAHB, A.; BELKHODJA, M.; MULET, J. M. 2016. Effect of salt stress on growth, chlorophyll content, lipid peroxidation and antioxidant defense systems in *Phaseolus vulgaris* L. South African Journal of Botany 105: 306-312.
- 49.- TAYLOR, A.; ALLEN, P.; BENNE-TT, M.; BRADFORD, K.; BURRIS, J.; MISRA, M. 1998. Seed enhancements. Seed Science Research 8(2): 245-256.
- 50.- THOMAS, U.C.; VARUGHESE, K.; THOMAS, A.; SADANANDAN, S. 2000. Seedpriming for increased vigor, viability and productivity of upland rice. Leisa India 4: 14.
- **51.- YOUSOF, A. F. I.** 2013. Effect of rice seed priming with calcium chloride on germination and seedlings vigor under salinity stress. Journal of Plant Production 4 (4): 523-535.